

# GENKWANIN 4'-GALACTOSIDE AND OTHER CONSTITUENTS FROM *DUABANGA SONNERATIOIDES*\*

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**Key Word Index**—*Duabanga sonneratioides*; Lythraceae.

Our initial investigations<sup>1,2</sup> on the stem bark of *D. sonneratioides* Ham. (Lythraceae) showed the presence of several known constituents. Further work on this plant, because of its activity against Walker's carcinosarcoma<sup>3</sup> has led to the isolation of ellagic acid, tetramethylellagic acid, epioleonic acid and a new flavone galactoside.

The flavone galactoside (**1**)  $C_{22}H_{22}O_{10}$ , m.p.  $260^{\circ}$  (dec.),  $\nu_{\max}$  3400, 1650, 1600, 1555, 1248, 1060 and  $830\text{ cm}^{-1}$ .  $\lambda_{\max}$  (EtOH) 226, 270 and 325 nm ( $AlCl_3$ ) 280, 300, 335 and 385 nm (NaOAc) 270 and 325 nm (NaOAc-boric acid) 226, 270 and 325 nm is sparingly soluble in  $CHCl_3$ ,  $Me_2CO$ , EtOAc and MeOH and gave positive Fiegels' test for glycoside. On acidic hydrolysis, **1** furnished an aglycone (**2**),  $C_{16}H_{12}O_5$ , m.p.  $267^{\circ}$ ,  $M^+$  284,  $\lambda_{\max}$  225, 270 and 329 nm (NaOAc) 225, 270 and 329 nm identical with genkwain<sup>4</sup> and a sugar, galactose (PC). **2** formed a diacetate,  $C_{20}H_{16}O_7$ , m.p.  $204^{\circ}$  (IR, NMR and MS).

The shift in the UV maxima of **1** in the presence of  $AlCl_3$  indicated the presence of a free 5-hydroxyl. This shows that the sugar moiety must be at position 7. It is known that glycosidation of the 4'-hydroxyl group produces a 3–10 nm hypsochromic shift in band 1.<sup>5</sup> Analogous with this,  $\lambda_{\max}$  329 nm of **2** shifts to 325 nm in **1**. The chemical and spectroscopic evidence thus indicated **1** to be 4'-O-galactoside of 5,4'-dihydroxy-7-methoxy-flavone.

Ellagic acid  $C_{14}H_6O_8$ , m.p.  $>360^{\circ}$ ,  $M^+$  302,  $\lambda_{\max}$  258 and 368 nm gave a positive Griessmayer test (IR and m.m.p.). It formed a tetraacetate  $C_{22}H_{14}O_{12}$ , m.p.  $336^{\circ}$  and a tetramethyl ether  $C_{18}H_{14}O_8$ , m.p.  $340^{\circ}$ . Tetramethyl ellagic acid  $C_{18}H_{14}O_8$ , m.p.  $340^{\circ}$  was characterized on the basis of IR, UV, MS and m.m.p.

Epioleonic acid  $C_{30}H_{48}O_3$ , m.p.  $302^{\circ}$  has been identified through its IR, MS and NMR and acetate  $C_{32}H_{50}O_4$ , m.p.  $268^{\circ}$ .

## EXPERIMENTAL

UV spectra were determined in EtOH, IR spectra in KBr and 60 MHz NMR spectra in  $CDCl_3$  with TMS as internal standard. Uncorrected capillary m.p.s are reported.

*Extraction and separation of constituents.* Air dried powdered stem bark (6 kg) were extracted with 50% EtOH

<sup>1</sup> BHAKUNI, D. S., GUPTA, N. C., SATISH, S., SHARMA, S. C., SHUKLA, Y. N. and TANDON, J. S. (1971) *Phytochemistry* **10**, 2247.

<sup>2</sup> SHARMA, S. C., SHUKLA, Y. N. and TANDON, J. S. (1972) *Phytochemistry* **11**, 2621.

<sup>3</sup> BHAKUNI, D. S., DHAR, M. L., DHAR, M. M., DHAWAN, B. N., GUPTA, B. and SRIMAL, R. C. (1971) *Ind. J. Exp. Bio.* **9**, 91.

<sup>4</sup> HASEGAWA, M. and SHIRATO, T. (1952) *J. Am. Chem. Soc.* **74**, 6114.

<sup>5</sup> MABRY, T. J., MARKHAM, K. R. and THOMAS, M. B. (1970) *The Systematic Identification of Flavonoids*, p. 45, Springer, Berlin.

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(40 l.) and the extract concentrated *in vacuo* below 50°. The syrupy material obtained was extracted successively with  $C_6H_6$  (6 l.), EtOAc (6 l.) and *n*-BuOH (4 l.).  $C_6H_6$  and EtOAc fraction gave compounds reported earlier<sup>1,2</sup> and also tetramethylellagic acid (10%, EtOAc in  $C_6H_6$ ) and epioleonic acid (20%, EtOAc in  $C_6H_6$ ). *n*-BuOH extract was freed of the solvent and the residue treated with  $Me_2CO$ .  $Me_2CO$  soluble fraction gave **1** while  $Me_2CO$  insoluble part afforded ellagic acid.

*Genkwanin 4'-galactoside (1)*.  $Me_2CO$  soluble part of the *n*-BuOH extract, deposited a solid which was recrystallized from EtOH (Found: C, 58.70; H, 5.20.  $C_{22}H_{22}O_{10}$  required: C, 59.10; H, 4.91%). Acid hydrolysis and extraction with  $Et_2O$  afforded genkwanin, m.p. 267° (3 crystallization gave m.p. 280°) (Found: C, 67.10; H, 4.30.  $C_{16}H_{12}O_5$  required: C, 67.60; H, 4.20%).

*Diacetate*. M.p. 204° (Found: C, 64.10; H, 4.10.  $C_{20}H_{16}O_7$  required: C, 64.60; H, 4.30%). The aq. hydrolysis soln was neutralized ( $BaCO_3$ ) and PC indicated that the only sugar present was galactose.

*Ellagic acid*. The  $Me_2CO$  insoluble fraction from the *n*-BuOH extract was extracted (Soxhlet) with hot  $Me_2CO$ . Concentration and standing in the cold furnished a yellow solid. This on recrystallization gave yellow needles, m.p. > 360° (Found: C, 56.10; H, 2.10.  $C_{14}H_6O_8$  required: C, 56.60; H, 1.98%).

*Tetramethyl ellagic acid*. Obtained as needles when crystallized from MeOH and dioxane m.p. 340° (Found: C, 59.80; H 4.20.  $C_{18}H_{14}O_8$  required: C, 60.33; H, 3.91%).

*Epioleonic acid*. Crystallized from MeOH, m.p. 302° (Found: C, 78.50; H, 9.86.  $C_{30}H_{48}O_3$  required: C, 78.94; H, 9.86%).

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## ACORENONE-B IN *ANGELICA LUCIDA* OIL

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*Plant*. *Angelica lucida* L. ( $\equiv$  *Coelopleurum actaeifolium* (Michx.) Coult. & Rose, *Archangelica gmelini* (specimen retained in Herbarium at University of Waterloo, NA 5422). *Source*. Blue Rocks, Lunenburg, Nova Scotia, Canada. *Uses*. Tender parts of plant used as food and tonic<sup>1</sup> by Eskimos.

*Present work*. The dried seeds were water distilled to yield 0.15% essential oil. The oil, which was analysed by a combination of techniques described previously<sup>2</sup> was found to contain acorenone-B (43.1%),  $\beta$ -phellandrene (16.1%), myrcene (8.0%), caryophyllene (7.5%),  $\alpha$ -pinene (2.0%), phenyl ethyl isovalerate (2.0%), bornyl acetate (1.6%), benzyl isovalerate (1.1%), camphene (0.3%),  $\beta$ -pinene (0.2%) and sabinene (0.2%).

*Comment*. Acorenone B, which was recently isolated from *Bothriochloa intermedia* (R.Br.) A. Camus (Gramineae) and synthesized,<sup>3</sup> was found to be identical to acorenone previously isolated from *Acorus calamus* L. (Araceae).<sup>4</sup>

<sup>1</sup> FRENCH, D. H. (1971) *The Biology & Chemistry of the Umbelliferae* (HEYWOOD, V. H., ed.), pp. 385–412, Publ. for Linn. Soc., Academic Press, London.

<sup>2</sup> LAWRENCE, B. M. (1971) *Can. Inst. Food Technol. J.* **4**, A44.

<sup>3</sup> McCCLURE, R. J. (1969) Ph.D. Thesis, Georgia Inst. Technol.

<sup>4</sup> VRKOC, J., HEROUT, V. and SORM, F. (1961) *Coll. Czech. Chem. Commun.* **26**, 3183.